

An efficient stereoselective synthesis of (2*S*,4*S*,5*R*)-(–)- and (2*R*,4*R*,5*S*)-(+)-bulgecinine

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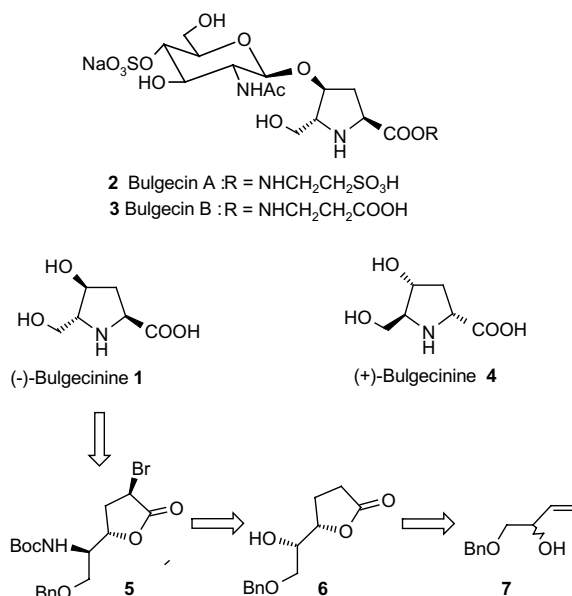
Abstract—A short synthetic route to (–)- and (+)-bulgecinine, the amino acid moiety of the bulgecins was achieved from the readily available nonchiral pool starting material *cis*-2-butene-1,4-diol in which a Claisen orthoester rearrangement and a Sharpless asymmetric dihydroxylation were used as the key steps.

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Bulgecinine **1** is an amino acid constituent of naturally occurring antibiotic glycopeptides called bulgecins (**2**, **3**), isolated from *Pseudomonas acidophila* and *Pseudomonas mesoacidophila*¹ bulgecins although devoid of antibacterial activity, induce characteristic morphological changes, called bulge formation, in the cell wall of Gram-negative bacteria in co-operation with β -lactam antibiotics. As a result of bulge formation, the activity of these antibiotics is effectively enhanced and the bacteria are killed at lower β -lactam concentrations. The structure of (–)-bulgecinine **1** has been determined chemically and crystallographically to be (2*S*,4*S*,5*R*)-4-hydroxy-5-hydroxymethyl proline.²

As a consequence of their biological effects³ and structural novelty, several total syntheses of (–)-bulgecinine have been reported in the literature.⁴ Herein we describe our strategy involving a Sharpless asymmetric dihydroxylation approach for the synthesis of (–)- and (+)-bulgecinine. In connection with our current studies directed towards the construction of biologically active compounds from the versatile intermediate **6**,⁵ we focused our attention on the stereoselective synthesis of (–)- and (+)-bulgecinine starting from the readily available nonchiral pool starting material *cis*-2-butene-1,4-diol.

The retrosynthetic analysis for our planned synthesis of (–)-bulgecinine is shown in Scheme 1. The bromolactone **5**, the precursor to bulgecinine, could be obtained

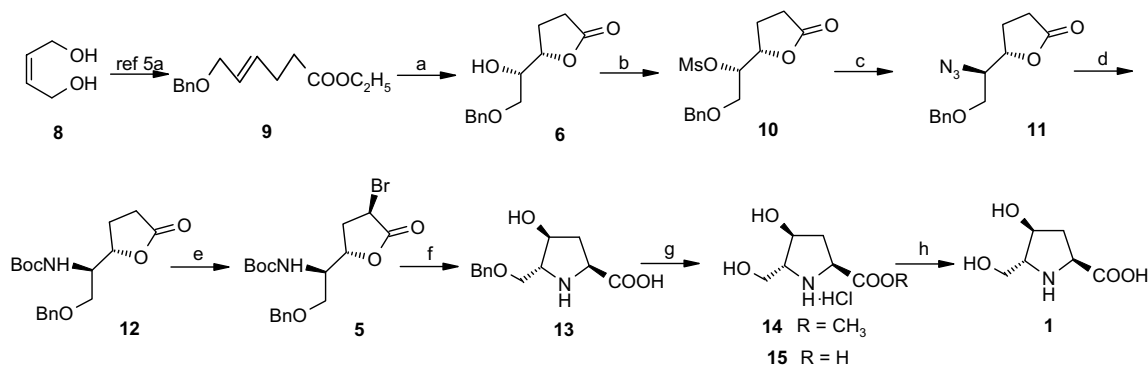


Scheme 1. Retrosynthetic analysis of **1**.

from the hydroxylactone **6**, which in turn was obtained from the allyl alcohol **7**.

As outlined in Scheme 2, the enantiomerically pure hydroxylactone **6**, was obtained in four steps from *cis*-2-butene-1,4-diol **8**, in which a Claisen orthoester rearrangement and a Sharpless asymmetric dihydroxylation were used as the key steps to install the requisite chirality. Mesylation of the hydroxylactone **6** using MsCl and

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Scheme 2. Reagents and conditions: (a) AD-mix- α , $\text{CH}_3\text{SO}_2\text{NH}_2$, $t\text{-BuOH-H}_2\text{O}$ (1/1), 24 h, 0 °C, 95%, 93% ee; (b) $\text{CH}_3\text{SO}_2\text{Cl}$, Et_3N , DMAP, DCM, 0 °C, 4 h, 92%; (c) NaN_3 , DMF, 90 °C, 24 h, 89%; (d) H_2 , 10% Pd-C, Et_3N , Boc_2O , EtOAc , rt, 2 h, 88%; (e) LiHMDS, TMSCl, Et_3N , NBS, THF, -78 °C, 2 h, 65%; (f) (i) CF_3COOH , DCM, 60 °C, 3 h, (ii) 0.1 N Ba(OH)_2 , pH = 9, 3 h, (iii) dil HCl, Amberlite-IR-120, 24 h, (iv) 6 N NH_4OH (aq), 3 h; (g) 10% Pd-C, methanolic HCl, rt, 24 h; (h) 1 M NaOH, rt, 92% (over three steps).

Et_3N in the presence of a catalytic amount of DMAP gave the mesylated lactone **10**. Displacement of the mesylate with NaN_3 at 90 °C in DMF gave the azidolactone **11**. Catalytic hydrogenation of the azido lactone over 10% Pd-C in ethyl acetate in the presence of Boc_2O gave the Boc protected lactone **12**.

The challenging task in our synthesis was to introduce the bromine stereoselectively at C-2 of the lactone **12**. This was achieved using LiHMDS, Et_3N and NBS at -78 °C for 2 h. The major isomer **5** along with its diastereomer (9:1 ratio) was separated by silica gel column chromatography. The conditions developed by Oppolzer⁶ were adapted to convert the major isomer (2*R*,4*S*,5*R*)-**5** into the (2*S*,4*S*,5*R*)-**13** using the following sequence of reactions. The lactone **5** was refluxed in DCM with 10 molequiv of CF_3COOH for 3 h followed by evaporation of the solvent. The residue was then dissolved in 0.1 N Ba(OH)_2 and the mixture maintained at pH 9 for 3 h. Acidification to pH 1, stirring the aqueous solution with Amberlite-IR-120 ion-exchange resin for 24 h, filtration, washing the resin with distilled water (until it showed clear solution with AgNO_3 to test the complete removal of Cl^-), stirring the resin with 6 N aqueous NH_4OH solution for 3 h and filtration gave the benzyl protected (2*S*,4*S*,5*R*)-(-)-bulgecinine **13**. Removal of the benzyl protection was performed using Pd/C in methanolic HCl under normal hydrogen pressure and at room temperature to deliver a mixture of (-)-bulgecinine hydrochloride **15** and its methyl ester **14**. This mixture was subjected to basic hydrolysis using 1 M NaOH to deliver (-)-bulgecinine **1**.

Accordingly we also prepared (+)-bulgecinine **4** following the same sequence of reactions from the opposite enantiomer of hydroxylactone **6**, which was prepared from the γ,δ -unsaturated ester **9** using AD-mix- β . The physical and spectroscopic data of all the synthetic intermediates were in good agreement with the proposed structures and those of **1** and **4** are in good agreement with the literature data.⁷

In summary (+)-and (-)-bulgecinine were synthesized in an efficient overall yield of 43% in seven steps from the

hydroxylactone **6**, which in turn was obtained from the readily available *cis*-2-butene-1,4-diol, as the common achiral precursor. The syntheses of other biologically active compounds from the versatile intermediate **6** are being investigated in our laboratory.

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References and notes

- (a) Imada, A.; Kintaka, K.; Nakao, M.; Shinagawa, S. *J. Antibiot.* **1982**, *35*, 1400–1403; (b) Shinagawa, S.; Maki, M.; Kintaka, K.; Imada, A.; Asai, M. *J. Antibiot.* **1985**, *38*, 17–23.
- Shinagawa, S.; Kashara, F.; Wada, Y.; Harada, S.; Asai, M. *Tetrahedron* **1984**, *40*, 3465–3470.
- For biological studies see (a) van Asselt, E. J.; Kalk, K. H.; Dijkstra, B. W. *Biochemistry* **2000**, *39*, 1924–1934; (b) Thunnissen, A. M. W. H.; Rozeboom, H. J.; Kalk, K. H.; Dijkstra, B. W. *Biochemistry* **1995**, *34*, 12729–12737; (c) Templin, M. F.; Edwards, D. H.; Hoeltje, J. Y. *J. Biol. Chem.* **1992**, *267*, 20039–20043, and references therein.
- For earlier syntheses of (-)-bulgecinine, see (a) Kharaf, J. K.; Datta, A. *J. Org. Chem.* **2004**, *69*, 387–390; (b) Holt, K. A.; Swift, J. P.; Smith, M. E. B.; Taylor, S. J. C.; McCague, R. *Tetrahedron Lett.* **2002**, *43*, 1545–1548; (c) Krasinski, A.; Jurczak, J. *Tetrahedron Lett.* **2001**, *42*, 2019–2021; (d) Burk, M. J.; Allen, J. G.; Kiesman, W. F. *J. Am. Chem. Soc.* **1998**, *120*, 657–663; (e) Maeda, M.; Okazaki, F.; Murayama, M.; Tachibana, Y.; Aoyagi, Y.; Ohta, A. *Chem. Pharm. Bull.* **1997**, *45*, 962–965; (f) Mazon, A.; Najera, C.; Ezquerro, J.; Pedregal, C. *Tetrahedron Lett.* **1997**, *38*, 2167–2170; (g) Panday, S. K.; Langlois, N. *Synth. Commun.* **1997**, *27*, 1373–1384; (h) Fehn, S.; Burger, K. *Tetrahedron: Asymmetry* **1997**, *8*, 2001–2005; (i) Graziani, L.; Porzi, G.; Sandri, S. *Tetrahedron: Asymmetry* **1996**, *7*, 1341–1346; (j) Yuasa, Y.; Ando, J.; Shibuya, S. *J. Chem. Soc., Perkin Trans. 1* **1996**, 793–802; (k) Madau, A.; Porzi, G.; Sandri, S. *Tetrahedron: Asymmetry* **1996**, *7*, 825–830; (l) Schmeck, C.; Hegedus, L. S. *J. Am. Chem. Soc.* **1994**, *116*, 9927–9934; (m) Jackson,

- R. F. W.; Rettie, A. B.; Wood, A.; Wythes, M. J. *J. Chem. Soc., Perkin Trans. 1* **1994**, 1719–1726; (n) Yuasa, Y.; Ando, J.; Shibuya, S. *J. Chem. Soc., Chem. Commun.* **1994**, 1, 1383–1384; (o) Jackson, R. F. W.; Rettie, A. B. *Tetrahedron Lett.* **1993**, 34, 2985–2986; (p) Hirai, Y.; Terada, T.; Amemiya, Y.; Momose, T. *Tetrahedron Lett.* **1992**, 33, 7893–7894; (q) Barrett, A. G. M.; Pilipauskas, D. *J. Org. Chem.* **1991**, 56, 2787–2800; (r) Barrett, A. G. M.; Pilipauskas, D. *J. Org. Chem.* **1990**, 55, 5194–5196; (q) Ohta, T.; Hosoi, A.; Nozoe, S. *Tetrahedron Lett.* **1988**, 29, 329–332; (t) Bashyal, B. P.; Chow, H. F.; Fleet, G. W. J. *Tetrahedron* **1987**, 43, 423–430; (u) Ofune, Y.; Hori, K.; Sakaitani, M. *Tetrahedron Lett.* **1986**, 27, 6079–6082; (v) Bashyal, B. P.; Chow, H. F.; Fleet, G. W. J. *Tetrahedron Lett.* **1986**, 27, 3205–3208; (w) Wakamiya, T.; Yamanoi, K.; Nishikawa, M.; Shiba, T. *Tetrahedron Lett.* **1985**, 26, 4759–4760.
5. (a) Chavan, S. P.; Praveen, C. *Tetrahedron Lett.* **2004**, 45, 421–423; (b) Chavan, S. P.; Praveen, C.; Ramakrishna, G.; Kalkote, U. R. *Tetrahedron Lett.* **2004**, 45, 6028–6077.
 6. Oppolzer, W.; Moretti, R.; Zhou, C. *Helv. Chim. Acta* **1994**, 77, 2363–2380.
 7. All new compounds were characterized and gave satisfactory spectral data. Compound **6**: $[\alpha]_D^{24} + 40.59$ (c 1, CHCl₃). ¹H NMR (200 MHz, CDCl₃) δ , ppm: 2.25 (2H, m), 2.48 (2H, m), 2.69 (1H, m), 3.59 (2H, m), 3.84 (1H, m), 4.57 (3H, m), 7.33 (5H, m). ¹³C NMR (50 MHz) δ : 23.3, 28.1, 70.5, 71.6, 73.1, 79.9, 127.5, 128.1, 137.5, 177.7 ppm. Anal. Calcd for C₁₃H₁₆O₄: C, 66.09; H, 6.83%. Found: C, 65.99; H, 6.77%. Compound **10**: $[\alpha]_D^{24} + 8.31$ (c 1, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ , ppm: 2.26 (1H, m), 2.36 (1H, m), 2.46 (1H, ddd, $J = 16.9, 10.1, 6.4$ Hz), 2.67 (1H, ddd, $J = 16.9, 9.6, 6.4$ Hz), 3.06 (3H, s), 3.71 (1H, dd, $J = 10.5, 3.6$ Hz), 3.83 (1H, dd, $J = 10.5, 7.3$ Hz), 4.55 (2H, m), 4.68 (1H, ddd, $J = 8.2, 5.5, 3.2$ Hz), 4.80 (1H, m), 7.32 (5H, m). ¹³C NMR (125 MHz) δ : 23.5, 27.3, 38.6, 66.9, 73.3, 77.5, 81.7, 127.7, 128.3, 136.9, 176.0 ppm. Anal. Calcd for C₁₄H₁₈O₆S: C, 53.49; H, 5.77; S, 10.2%. Found: C, 53.62; H, 5.86; S, 10.38%. Compound **11**: $[\alpha]_D^{24} - 15.83$ (c 1, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ , ppm: 2.12 (1H, m), 2.23 (1H, m), 2.44–2.59 (2H, m), 3.62 (1H, dd, $J = 10.1, 6.4$ Hz), 3.66 (1H, dd, $J = 10.1, 4.6$ Hz), 3.79 (1H, m), 4.55 (3H, m), 7.32 (5H, m). ¹³C NMR (125 MHz) δ : 23.1, 27.6, 63.0, 68.9, 73.1, 77.6, 127.3, 128.2, 137.1, 175.7 ppm. Anal. Calcd for C₁₃H₁₅NO₃: C, 59.77; H, 5.76; N, 16.08%. Found: C, 59.57; H, 5.84; N, 16.19%. Compound **12**: $[\alpha]_D^{24} + 1.87$ (c 1, CHCl₃). ¹H NMR (200 MHz, CDCl₃) δ , ppm: 1.39 (9H, s), 2.15 (2H, m), 2.43 (2H, m), 3.50 (1H, dd, $J = 9.4, 3.5$ Hz), 3.68–3.80 (2H, m), 4.45–4.56 (3H, m), 5.10 (1H, d, $J = 9.3$ Hz), 7.28 (5H, m). ¹³C NMR (125 MHz) δ : 24.4, 27.8, 28.1, 52.9, 68.8, 73.3, 78.5, 79.6, 127.5, 128.2, 137.5, 155.3, 176.2 ppm. Anal. Calcd for C₁₈H₂₅NO₅: C, 64.46; H, 7.51; N, 4.17%. Found: C, 64.38; H, 7.32; N, 4.08%. Compound **5**: $[\alpha]_D^{24} + 10.50$ (c 1, CHCl₃). ¹H NMR (200 MHz, CDCl₃) δ , ppm: 1.36 (9H, s), 2.42 (1H, ddd, $J = 8.2, 5.9, 2.3$ Hz), 2.68 (1H, m), 3.52 (1H, dd, $J = 9.8, 3.5$ Hz), 3.80 (2H, m), 4.42 (1H, dd, $J = 6.6, 2.3$ Hz), 4.51 (2H, m), 4.78 (1H, m), 5.10 (1H, d, $J = 9$ Hz), 7.26 (5H, m). ¹³C NMR (125 MHz) δ : 28.3, 36.8, 38.3, 52.8, 68.7, 73.5, 77.5, 80.0, 127.7, 128.4, 137.4, 155.3, 171.6 ppm. Anal. Calcd for C₁₈H₂₄NO₅Br: C, 52.18; H, 5.83; N, 3.37; Br, 19.28%. Found: C, 52.07; H, 5.97; N, 3.36; Br, 19.06%. Compound **13**: $[\alpha]_D^{24} - 8.09$ (c 1, H₂O). ¹H NMR (500 MHz, D₂O) δ , ppm: 2.11 (1H, ddd, $J = 14.1, 6.2, 4.3$ Hz), 2.51 (1H, ddd, $J = 14.1, 9.0, 5.8$ Hz), 3.56–3.71 (3H, m), 4.06 (1H, dd, $J = 9.0, 6.6$ Hz), 4.23 (1H, m), 4.49 (2H, m), 7.32 (5H, m). ¹³C NMR (125 MHz) δ : 36.2, 59.1, 64.8, 65.7, 70.6, 72.8, 128.0, 128.4, 136.7, 173.6 ppm. Anal. Calcd for C₁₃H₁₇NO₄: C, 62.14; H, 6.81; N, 5.57%. Found: C, 62.02; H, 6.92; N, 5.49%. Compound **1**: $[\alpha]_D^{24} - 13.3$ (c 0.9, H₂O). ¹H NMR (200 MHz, D₂O) δ , ppm: 2.11 (1H, ddd, $J = 14.5, 6.3, 4.6$ Hz), 2.58 (1H, ddd, $J = 14.5, 9.0, 5.9$ Hz), 3.56–3.71 (3H, m), 4.06 (1H, dd, $J = 9.0, 6.6$ Hz), 4.34 (1H, m), ¹³C NMR (75 MHz) δ : 37.2, 59.2, 60.5, 66.2, 71.7, 174.7 ppm.